

E18-2014-11

P. S. Nekhoroshkov*, A. V. Kravtsova*,
M. V. Frontasyeva, Yu. N. Tokarev*

NEUTRON ACTIVATION ANALYSIS
AND SCANNING ELECTRON MICROSCOPY
OF PHYTOPLANKTON IN THE COASTAL ZONE
OF THE CRIMEA (BLACK SEA)

Submitted to «American Journal of Analytical Chemistry»

*The A. O. Kovalevsky Institute of Biology of the Southern Seas of the
National Academy of Sciences of Ukraine, Sevastopol, Ukraine

Нехорошков П. С. и др.

E18-2014-11

Нейтронный активационный анализ и сканирующая электронная микроскопия фитопланктона прибрежной зоны Крыма (Черное море)

Впервые для прибрежной зоны Черного моря (Севастопольский регион, Украина) с применением нейтронного активационного анализа и сканирующей электронной микроскопии с энергодисперсионной рентгеновской спектроскопией определены концентрации 45 элементов в планктонных сообществах, используемых в качестве биоиндикаторов неорганического загрязнения прибрежных вод. Образцы фитопланктона были отобраны планктонной сетью (диаметр пор 35 мкм) путем тотальных ловов на 3 станциях, расположенных в загрязненных и условно чистых акваториях прибрежной зоны Севастополя, в течение осеннего периода активного роста фитопланктона. Концентрации Mg, Al, Sc, Ti, V, Mn, As, Rb, Ba, Th и Fe, Cr возрастают экспоненциально от условно чистой станции к более загрязненной, и в 10 и 3 раза выше, соответственно, в фитопланктоне Севастопольской бухты. Концентрации редкоземельных элементов на всех станциях примерно одинаковы, менее 1 мкг/г, при этом имеют тенденцию накапливаться в фитопланктоне загрязненной станции в Севастопольской бухте. Полученные результаты хорошо согласуются с данными о концентрациях элементов в океаническом планктоне, планктонных сообществах Белого и Черного морей. С использованием энергодисперсионной рентгеновской спектроскопии в фитопланктоне на загрязненной станции были обнаружены минеральные частицы неизвестного происхождения с включениями меди (0,42 % по весу), а на условно чистой — с включениями цинка (0,57 % по весу).

Работа выполнена в Лаборатории нейтронной физики им. И. М. Франка ОИЯИ.
Препринт Объединенного института ядерных исследований. Дубна, 2014

Nekhoroshkov P. S. et al.

E18-2014-11

Neutron Activation Analysis and Scanning Electron Microscopy of Phytoplankton in the Coastal Zone of the Crimea (Black Sea)

For the first time the concentrations of 45 elements in the coastal phytoplankton communities used as bioindicator of inorganic contamination of the Black Sea coastal area near Sevastopol, Ukraine, were determined by means of neutron activation analysis and scanning electron microscopy with energy dispersive spectrometer. Phytoplankton samples were collected by total tows of the plankton net with 35 μm pore size at 3 stations situated in polluted and relatively pristine water areas of the Sevastopol coastal zone during autumn period of the phytoplankton active growth. The concentration of Mg, Al, Sc, Ti, V, Mn, As, Rb, Ba, Th and Fe, Cr increases exponentially from relatively pristine station to more polluted station and 10 and 3 times greater, respectively, in the phytoplankton of the Sevastopol Bay. The rare-earth elements have relatively the same concentration values of about 1 $\mu\text{g/g}$ and tend to accumulate in the phytoplankton from the polluted station in the Sevastopol Bay. The obtained results are in good agreement with the elemental concentration data in the oceanic plankton, plankton communities from the White Sea and the Black Sea. Using energy-dispersive X-ray spectrometry the mineral particles of unknown origin and impurities of copper (0.42 % by weight) in the phytoplankton at the polluted station and zinc (0.57 % by weight) at the relatively pristine station were determined.

The investigation has been performed at the Frank Laboratory of Neutron Physics, JINR.
Preprint of the Joint Institute for Nuclear Research. Dubna, 2014

INTRODUCTION

Plankton (phytoplankton and zooplankton) containing organisms suspended in water and typically carried by the waves, tides and currents is very sensitive to changes in the environment; therefore, it is widely used as an indicator of water quality [1–3]. Elemental accumulation as a dynamic process in plankton depends on concentration of elements in the environment and different processes induced by the environmental nutritional stress, which are determined by functional state of each organism [4]. Earlier it was shown that the plankton responds to the presence of heavy metals in water environment, upon that some elements inhibit and some stimulate growth of organisms and community [1, 2].

The physiology and ecology of planktonic organisms are influenced by the concentration, chemical speciation and resulting bioavailability of some trace metals such as Fe, Cu, Zn, Cd, and Hg. Therefore, since A.P. Vinogradov [5] determined the concentrations of Ca, Mg, K, P, Fe, Mn, S, Cl, Si, Br, and I in zooplankton sampled in the White and Barents Seas, many studies have been devoted to chemical (elemental) content of the plankton in the different regions of the World Ocean.

Goals and approaches to the study of the elemental concentration of plankton are very different. Thus, Leonova estimated the concentration of a wide range of elements in the White Sea zooplankton [6], phytoplankton and zooplankton from freshwater Siberian lakes [7] and considered their accumulation capacity and involving into geochemical processes. The ability of plankton to concentrate trace elements from its environment is used to indicate the level of coastal waters pollution in different regions of the World Ocean [2, 8–12].

The elemental content of the Black Sea plankton has been studied for over 50 years. Thus, Vinogradova [13] studied the ability of plankton to accumulate polyvalent metals, Rozhanskaya [14] explored the distribution of Mn, Fe, Co, Ni, Cu, and Zn over the food chain from plankton to the fishes, Saenko [15] gave the data about the concentration of Ti, V, Cr, Mn, Fe, and Cu in the plankton

from the coastal waters of the Sevastopol region. But generally large zooplankton (more than 100 microns) and casual small amounts of phytoplankton was studied by the mentioned scientists and only up to 10 elements were determined.

Marine coastal area of the Sevastopol region is exposed to various types of pollution. Heavy metals and other pollutants enter into the coastal waters with the surface and underwater runoff [16]. Heavy metals also come from old ships, industrial and agricultural facilities. The grain terminal in the Sevastopol Bay, where several large vessels dock every day, is one more source of the coastal waters pollution with trace elements (ballast waters, paint and plank materials, fuels). Secondary pollution takes place during the storm wave when buried fuel and oil are getting up from the bottom sediments to the surface [16].

Since the anthropogenic impact on the plankton communities increases, the present study is focused on the elemental composition of the phytoplankton assemblages from the polluted zone of the Sevastopol Bay and from relatively pristine deep water coastal station.

To determine structure and elemental composition of phytoplankton and suspended particles, the methods of scanning electron microscopy (SEM), energy-dispersive X-ray spectrometry (EDS) and instrumental neutron activation analysis (INAA) were used.

The multielement INAA at the reactor IBR-2 of FLNP, JINR with its low temperature in the irradiation channels is particularly efficient in the analysis of biological samples [17]. The SEM/EDS also is widely applied in biological analysis [18]. To our knowledge, these two complementary techniques were never simultaneously applied to study the Black Sea phytoplankton.

MATERIALS AND METHODS

Sampling and Sample Preparation. The samples of phytoplankton were collected in the autumn of 2013 during its active growth at three coastal stations, where the first and the second stations are located in a relatively pristine zone, and the third one in the polluted zone of the Sevastopol Bay (Fig. 1). Stations 1 and 2 are situated in deep waters (the depth is more than 40 meters) and station 3 is in the entrance of the Sevastopol Bay (12 m depth). The characteristics of the sampling sites are shown in Table 1.

The phytoplankton was collected by means of the vertical tows of the phytoplankton Nansen net (35 μm mesh size) [19].

Sampling sites were chosen in compliance with the data of hydrochemical monitoring and the quality of water according to the trophical index (TRIX) of the coastal waters in the Sevastopol region [20]. Samples were filtered on paper filters with 10 μm pore size in the laboratory conditions. The sea-salt

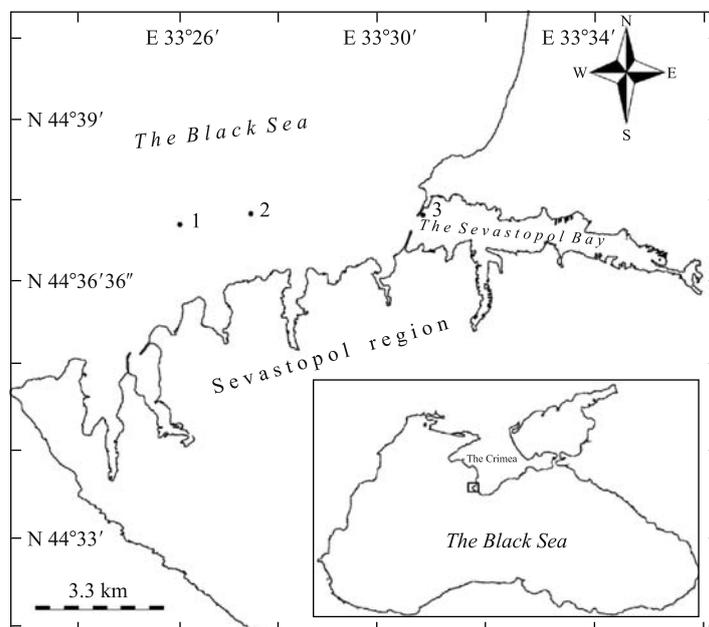


Fig. 1. Sampling sites in the coastal zone of the Sevastopol region (Crimea, the Black Sea)

Table 1. Sampling station

Number of station	Coordinates		Maximal depth, m	Distance from the shore, m	Filtered water volume, m ³
1	44°37'51.6" n. l.	33°26'20.2" e. l.	60	2660	10.75
2	44°37'45.01" n. l.	33°27'22.46" e. l.	45	2010	8.60
3	44°37'30.21" n. l.	33°30'41.51" e. l.	12	141	3.22

particles were dissolved and washed out after the treatment with distilled water. Then the filters with phytoplankton were dried at room temperature and subdivided for INAA and SEM/EDS.

Analysis. Elemental analysis of the phytoplankton samples was carried out by INAA at the reactor IBR-2 of FLNP, JINR. The analytical procedures and the basic characteristics of the employed experimental facility are described in detail elsewhere [17]. The samples of about 0.3 g were packed in polyethylene bags for short-term irradiation and in aluminum cups for long-term irradiation.

To determine the short-lived isotopes of Mg, Al, Cl, Ca, Ti, V, Mn, Cu, and I, the samples were irradiated for 3 min in the reactor channel with a neutron flux density of $1.3 \cdot 10^{12} \text{ n}/(\text{cm}^2 \cdot \text{s})$. Gamma spectra of induced activity were measured for 12–15 min after 20 min of decay. The elemental contents of the long-lived isotopes of Na, K, Sc, Cr, Fe, Co, Ni, Zn, As, Se, Br, Rb, Sr, Zr, Mo, Ag, Sb, Cs, Ba, La, Ce, Nd, Sm, Gd, Tb, Tm, Yb, Hf, Ta, W, Au, Th, and U were determined using epithermal neutrons in a cadmium-screened irradiation channel with a neutron flux density of $1.6 \cdot 10^{12} \text{ n}/(\text{cm}^2 \cdot \text{s})$. Samples were irradiated for 90 h, repacked and then measured twice after 4–5 d of decay during 30 minutes and after 20 days of decay during 1.5 hours.

To process gamma spectra of induced activity and to calculate concentrations of elements in the samples, software developed at FLNP, JINR was used [21]. The uncertainties in the determined concentrations were in the range of 5–15 %, and of 30 % or more for those elements which concentrations in the samples were at the detection limit, e. g., Nd, Tm, Yb.

Quality control was provided by using NIST standard reference materials: 433, 690CC, 1547, 1572, 1632b, 1633b, 2709, 2710 irradiated in the same conditions together with the samples under investigation.

The SEM/EDS of the phytoplankton specimens were performed using SMA Quanta 3D FEG instrument in the Laboratory of microscopy of Skolkovo Institute of Science and Technology. The SEM images of selected area of phytoplankton and spots were obtained simultaneously with the rapid quantitative analysis of the elemental concentrations.

RESULTS AND DISCUSSION

The concentrations of 45 elements determined by INAA in phytoplankton of the coastal zone of the Black Sea (stations 1–3), as well as comparative literature are given in Table 2.

The elemental concentrations in the phytoplankton were calculated for the coastal stations in relatively pristine water area (station 1), in relatively polluted water area (station 3 in the Sevastopol Bay) and in the intermediate zone (station 2) (Table 2). Our data were compared with the elemental concentrations in phytoplankton from the coastal waters of different regions of the World Ocean [15, 22, 23].

The concentrations of some elements (Na, K, and Ca) in the plankton from the White Sea [22] exceed our results because after filtration their samples were not washed by distilled water.

The concentrations of macro and trace elements which are significantly accumulated in phytoplankton are shown in Fig. 2.

Table 2. Elemental concentrations in the plankton ($\mu\text{g/g}$)

Element	Phytoplankton			Total plankton		
	Present study (INAA), the Black Sea			The White Sea [22]	World Ocean [23]	The Black Sea [15]
	Station 1 (relatively pristine area)	Station 2	Station 3 (polluted area)			
Na	2025±52	715±19	1095±29	63000	33000	
Mg	173±6	546±19	2370±69		94000	
Al	< 309	951±26	4421± 118		62	
S	50.8±23.5	25.4±11.6	4.6±3.2		8300	
Cl	2650±233	1520±134	1520±135		4700	
K	1380±787	732±288	1810±455	13000	52000	
Ca	< 811	349±35	4379±386	6000	14000	
Sc	< 0.03	0.33±0.10	0.75±0.23	0.45	0.07	
Ti	15±4	64±10	291±25	400	10	45–201
V	0.20±0.02	1.4±0.1	6.6±0.3	3	3.5	6–15
Cr	5.16±1.15	11.86±1.24	15.76±1.51	620	1.8	30–69
Mn	3.0 ±0.2	6.0±0.4	28.4±1.5	80	20	3–63
Fe	1370±198	1580±173	4220±295	2460	160	3310–9670
Ni	0.43±0.02	0.28±0.02	0.81±0.03	0.9	0.4	
Co	0.6±0.1	15.7±1.1	15.1±1.2	3.5	1.4	
Cu	91.4±32.4	43.1±15.7	117±42	43	12	38–187
Zn	205±20	86±8	124±11	362	39	
As	0.11±0.01	0.59±0.04	1.8±0.1	13	15	
Se	0.26±0.06	< 0.13	0.04±0.01	0.4	0.063	
Br	59.1±7.1	28.2±3.4	64.7±7.7	2000	440	
Rb	0.20±0.12	2.7±0.5	7.4±1.4	7	1.8	
Sr	6.3±0.8	6.8±0.7	15.1±1.4	106	1100	
Zr	7.0±4.7	8.0±3.5	16.7±6.5	3.6	0.7	
Mo	0.2±0.07	0.19±0.09	0.24±0.08	0.2	0.39	
Ag	0.07±0.01	0.07±0.02	0.05±0.013		0.22	
Sn	21.3±2.6	1.3±0.4	1.8±0.5	1.2	0.29	
Sb	0.74±0.03	0.28±0.01	0.37±0.04	1.8	0.16	
I	1.2±0.2	2.2±0.4	22.0±4.0	139	1020	
Ba	33.6±8.9	< 9.9 ± 4.5	48.7±13.4	28	19	
Cs	0.012±0.01	0.18±0.01	0.47±0.02	0.6	0.072	
La	0.115±0.025	0.822±0.055	2.1±0.12	1.18	0.14	
Ce	0.6±0.3	1.2±0.5	3.7±1.1	2.6	0.23	

The end of Table 2

Element	Phytoplankton			Total plankton		
	Present study (INAA), the Black Sea			The White Sea [22]	World Ocean [23]	The Black Sea [15]
	Station 1 (relatively pristine area)	Station 2	Station 3 (polluted area)			
Nd	3.1±1	0.5±0.4	1.1±1.3			
Sm	0.6±0.2	0.7±0.2	0.6±0.2			
Gd	< 0.6	4.0±1.2	1.8±0.6			
Tb	< 0.01	0.02±0.01	0.06±0.02	0.022		
Dy	< 0.14	0.11±0.03	0.32±0.11			
Tm	0.04±0.01	0.02±0.01	0.16±0.02			
Yb	0.14±0.07	0.11±0.04	0.53±0.12	0.13		
Hf	< 0.02	0.19±0.06	0.34±0.1	0.11		
Ta	0.002±0.001	0.03±0.01	0.05±0.02	0.06		
W	0.3±0.1	0.2±0.1	0.3±0.1		0.035	
Au	0.004±0.001	0.002±0.001	0.002±0.001		0.008	
Th	< 0.03	0.30±0.01	0.70±0.02	0.28	0.1	
U	2/4±0.2	1.9±0.2	0.7±0.1		0.8	

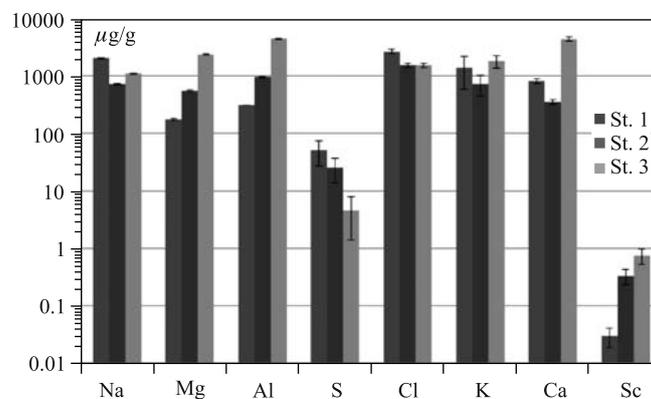


Fig. 2. Concentrations of major elements in phytoplankton ($\mu\text{g/g}$) determined by INAA at 3 stations (log scaling)

As follows from the results obtained, in phytoplankton the concentration of major elements from the most polluted shallow from water station 3 is considerable higher than in phytoplankton from the deeper stations. For example,

the concentration of such elements as Mg, Al, Sc, Ti, V, Mn, As, Rb, Ba, and Th (Figs. 2 and 3) exponentially increases towards more polluted station 3 by 5–10-fold. However, concentrations of Fe and Cr in the phytoplankton from the Sevastopol Bay are only 2-fold greater.

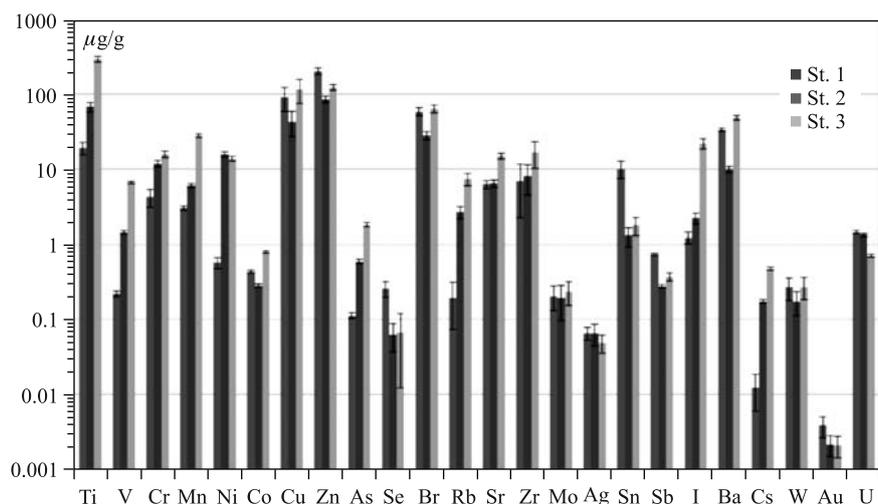


Fig. 3. Concentrations of some elements in the phytoplankton samples (log scaling)

The high content of polyvalent metals in the phytoplankton is due to the high concentrating ability of the phytoplankton. Thus, according to [15], concentrations of Mn, Fe, Ti and Cu, Cr in plankton (phyto- and zooplankton) collected in September 1969 in the Sevastopol Bay and seaward were 10-fold and 2–3-fold greater, respectively, than in the marine macrophytes collected at the same sampling site. Concentration of such elements as Na, Cl, K, Br, Mo, Ag, Sb, Au, and W is relatively constant in the phytoplankton from all three stations (Table 2). It is probably due to their homogeneous distribution in the coastal waters in this period. The content of iodine is constant in the phytoplankton from the deep water stations (1 and 2), but at the station 3 it is 10-fold greater. On the one hand, this is due to the presence of large number of macroalgae thalli (bodies) containing large amounts of I in the shallow water station. On the other hand, Vinogradov [5] notes the high content of iodine in diatoms that intensively grow in the shallow waters. The concentrations of the poorly mobile elements (Sc, Ti, V, Mn, Fe, Co, Ga, Rb, Y, Zr, Nb, Cs, Ba, lanthanides, Hf, Ta, and Th) are characteristic for the terrigenous component comprising 20–30% of the ash content in the seston, which is one order of magnitude higher in the seston [6]. A significant fraction of the terrigenous admixture was observed in the seston from the station 3.

It is explained a rather large difference (in the range of one order of magnitude) in values of elemental concentrations, indicators of the mineral sludge component of the suspension containing REE, Sc, Al, Ta, and Th [6].

The concentration values of the rare-earth elements are relatively of the same order (about 1 $\mu\text{g/g}$) (Fig.4), and they tend to accumulate in the phytoplankton from the polluted station in the Sevastopol Bay. This is probably due to a significant amount of disturbed bottom and terrigenous sediments in the phytoplankton samples.

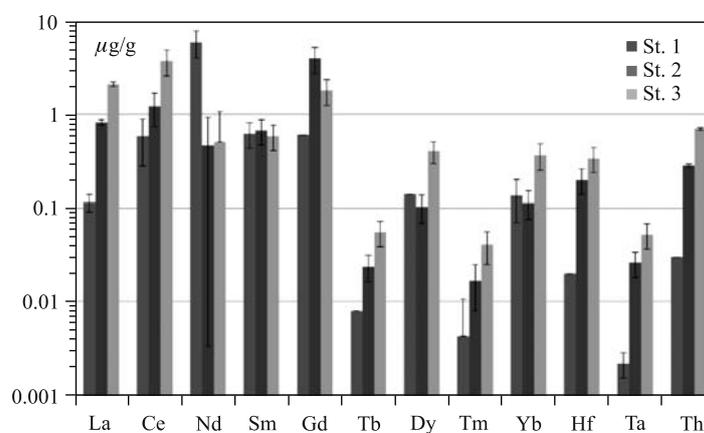


Fig. 4. Concentrations of the rare-earth elements in the phytoplankton from the 3 stations (log scaling)

Concentrations of the most elements in the phytoplankton at all stations are in good agreement with the concentrations in the total phyto- and zooplankton from the other regions of the World Ocean (with other salinity and other hydrological features) (Table 2).

The Black Sea phytoplankton community consists of many different species. The trophic structure, seasonal variability of phytoplankton community and quality of water (for example, transparency on disk Secchi depth) are mostly determined by the variability and dynamics of two groups of phytoplankton (diatoms and dinoflagellates). The diatoms (*Bacillariophyta*) are small autotrophic cells which consist of SiO_2 and which can live in the polluted coastal and estuarine waters in the photic zone. The phylum *Bacillariophyta* is distinguished by the presence of an inorganic cell-wall composed of silica (hydrated SiO_2).

The collected phytoplankton samples contain organic (both living phyto- and zooplankton and detrital), inorganic particles, and clay minerals; all together

called as seston [24]. Therefore, we should take into account that the SEM images show aggregates of mineral particles, parts and tissues of decaying indefinable organisms and living plankton cells (Figs. 5, 6, 9).

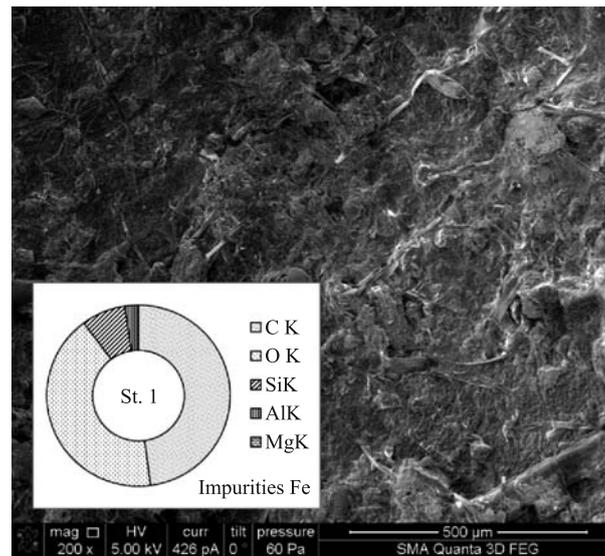


Fig. 5. The SEM image and elemental concentration (EDS) for full area of specimen from station 1

In the open coastal zone (stations 1 and 2) the dynamic water circulation is different in different seasons. In summer–autumn the relative biodiversity of phytoplankton community is greater in the deeper coastal water areas than at the stations located in the bay. Therefore, homotypical phytoplankton aggregation with only several species of diatoms grows in the Sevastopol Bay at a depth less than 15 m. SEM images of specimens from the shallow water from station 3 show that diatoms have line, curve and boxes shapes (Fig. 9). The number of dinoflagellates is less than a number of other species at this station, because dinoflagellates usually escaping polluted shallow water areas (e.g., Figs. 5, 6, 9).

Zooplankton in shallow waters (station 3) is practically absent, and at stations 1 and 2 in the deep water areas zooplankton organisms occur in samples with products of their living (fecal pellets, pieces of tissues).

Thus, a lot of diatoms phytoplankton algae of line and box shapes were observed in the specimen from the polluted station 3, but in the specimen from the relatively pristine deep water stations 1 and 2 mature adults of zooplankton and heterotrophic dinoflagellates were also found (Figs. 5 and 6). This fact shows

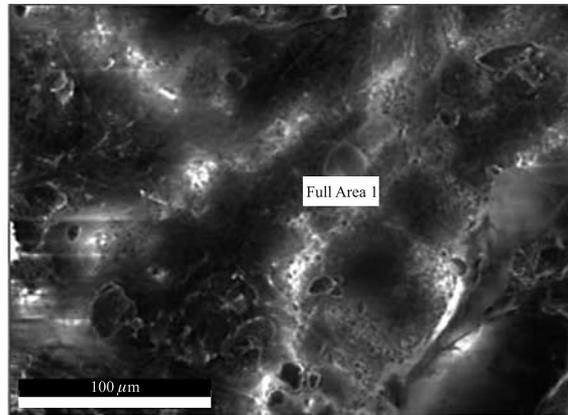


Fig. 6. SEM image of 300 μm phytoplankton specimen from station 2

that plankton (both phyto- and zooplankton) community in polluted shallow water coastal areas (station 3) has just a several number of species compared to relatively pristine deep water areas.

Organogenic elements such as C, O, N, H, P, and S compose biomass of living organisms. They form proteins, carbohydrates, lipids, nucleic acids and other biologically active compounds present in each cell of phytoplankton. Essential elements such as Ca, K, Na, Cl, Zn, Mn, Mo, I, Se, Mg, Fe, Cu, and Co are responsible for the physiological processes of cells [25]. However, high concentration of these elements in the environmental water can slow down the vital processes, such as photosynthesis in chloroplasts in the phytoplankton cells. Several diatom algae are covered with SiO_2 , and some elements might be embedded in their shells or tissues. Such elements have been found by EDS (Figs. 7, 8, 9).

The elemental concentrations on the surface of the specimen were estimated by the EDS in homogeneous material of each sample. The random selection of several spots in the field of specimen helps one to study a particulate elemental structure of the sample (Fig. 9).

All the studied specimens had relatively equal elemental concentration levels, both in full areas and randomly selected spots. Using SEM/EDS, we observed several spots with nontypical concentration features. For example, we observed spots without carbon (obviously of the mineral origin), with the oxides of Si, Al, Fe, and Mg; the calcium particles; biogenic particles (Figs. 8 and 9) with the Cu (0.42% by weight) and Zn (0.57% by weight) impurities of the unknown origin. The high concentrations of copper and zinc are likely associated with the anthropogenic pollution sources.

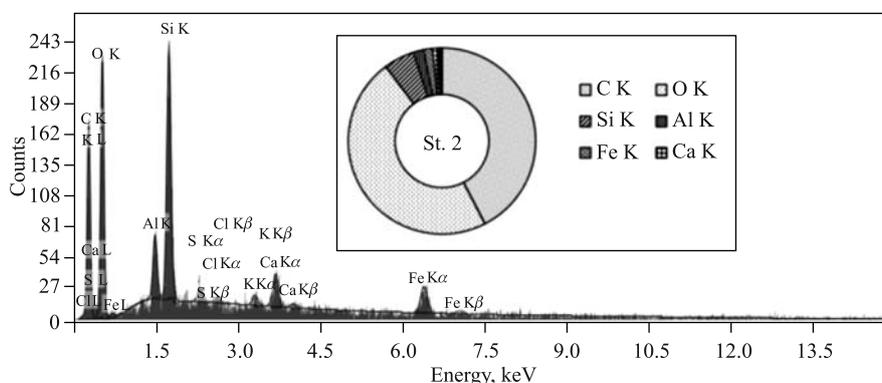


Fig. 7. X-ray spectrum of the 300 μm phytoplankton specimen from the station 2 (EDS)

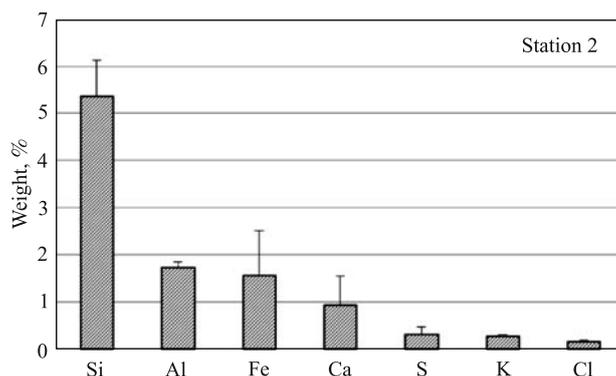


Fig. 8. Relative levels of the major element concentrations in the 300 μm phytoplankton specimen from the station 2 (EDS)

Ca, Al, Fe, Mg, K, Cl, and Na determined by INAA showed the highest concentrations in the plankton samples. Ca, Mg, K, Cl, and Na are the most abundant ions of seawater, and by weight these ions constitute $\sim 99\%$ of sea salts. High levels of Si and Al, which were detected by the EDS and INAA, confirm hypothesis that the silicon and aluminum cycles in the oceans are linked through the activity of diatoms [26]. Iron is undoubtedly the most versatile and important trace element for biochemical catalysis. The particulate and colloidal fraction comprises oxides, aluminosilicates and possibly organic forms [27].

The differences between concentrations of the major elements, which were determined by different methods, are associated with properties and features of

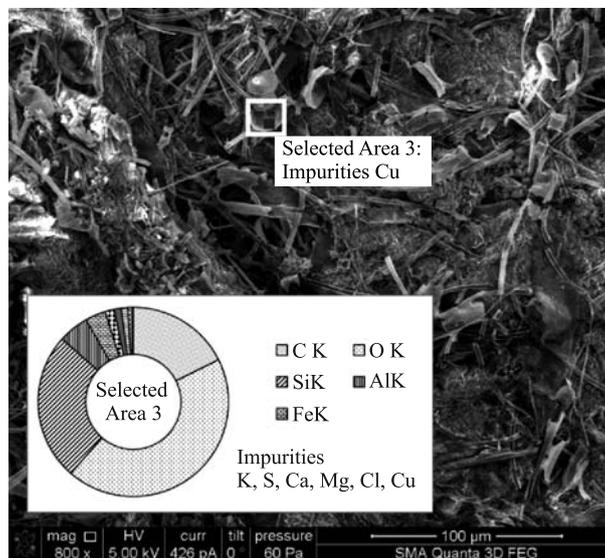


Fig. 9. 300 μm phytoplankton specimen from the station 3 and relative concentration levels for the selected area 3 with marked impurities (EDS)

the EDS and INAA. The detection limits of the elements determined by EDS are about 30–3000 ppm ($> 1\%$ by weight) as they depend on the energy and spatial resolution of the apparatus and they are greater compared to the detection limits and accuracy of INAA. However, elemental concentrations in the small patches and spots in the surface of the specimen were determined by the EDS. Therefore, the EDS is a rough and ready tool for the identification of the elements and analysis of the spatial elemental structure.

CONCLUSIONS

1. The results obtained show that the concentrations of major elements in phytoplankton from the shallow water from station 3 is greater than that in phytoplankton from the deeper stations. It means that phytoplankton from the Sevastopol Bay is involved in geochemical processes of the coastal elemental accumulation. The spatial distribution of terrigenous particles may, therefore, reflect the accumulation areas for such aggregates, although, it could also derive from lateral injection of resuspended sediment from the nearby continental shelf.

2. The concentration of Mg, Al, Sc, Ti, V, Mn, As, Rb, I, Ba, Th and Fe, Cr increases exponentially from the relatively pristine station 1 to more

polluted station 3, and they are 10- and 3-fold greater in the phytoplankton of the Sevastopol Bay, respectively. These data justify the opinion about the specific geochemical condition of the Sevastopol Bay that is typical for the marginal areas and appears also in increased concentrations of biophilic elements of the Fe group.

3. Concentrations of Na, Cl, K, Br, Mo, Ag, Sb, Au, and W are relatively constant in the phytoplankton from all three stations due to their homogeneous distribution in the coastal waters. Elements such as Na, K, Br, Ag, Au are known to be trace elements concentrated by biological processes in plants [28]. However, elements such as Sc, Ti, V, Th, Mo, W and others are not known to be concentrated by biological processes, so some other source of association may be indicated. Physical adsorption and entrapment of colloidal mineral matter by the gelatinous matrix of the diatoms are possible.

4. The concentration of rare-earth elements is relatively the same (less than 1 $\mu\text{g/g}$), and it tends to accumulate in the phytoplankton from the polluted station in the Sevastopol Bay, most probably, due to the significant amount of disturbed bottom and terrigenous sediments in the phytoplankton samples.

5. The SEM/EDS allowed determination of mineral particles of the unknown origin and impurities of copper (0.42% by weight) at the shallow water from station 3 and zinc (0.57% by weight) at the deep station 1.

6. A lot of diatoms phytoplankton microalgae of line and box shapes (ruderal species) were observed in the specimen from the station 3 in polluted area, but in the specimen from relatively pristine deep water from stations 1 and 2 only mature adults of zooplankton and heterotrophic dinoflagellates were found.

Our results are in good agreement with the elemental concentrations in phytoplankton from the other regions of the World Ocean. Phytoplankton in the coastal area of the Sevastopol region is experiencing anthropogenic impact, which is reflected in the elemental composition of phytoplankton and species structure. With simple modifications to the sampling and analytical techniques described here, size-fractionated phytoplankton and particulate concentrations for a broad suite of metals can be determined in a range of oceanic regimes, from productive coastal and estuarine environments to oligotrophic offshore waters.

Estimation of the elemental concentration levels in phytoplankton is an important part of the studying and understanding the pollution processes, because it shows ambient side of the human activity and the environmental influence on the state of the relatively close community of phytoplankton. The study of the elemental concentrations in phytoplankton assemblages with the estimation of their functional role in the polluted coastal areas will be continued.

This study is a part of the complex research of the effect of pollution on the phytoplankton in the coastal waters of the Crimean peninsula in the Sevastopol region and it was carried out in the framework of the monitoring program of the Black Sea coastal areas by the A.O. Kovalevsky Institute of Biology of the Southern Seas (IBSS). The findings of this study may serve as a background for

a future complex environmental monitoring of the coastal waters of the Crimea, where the data on concentrations of the elements and the biophysical parameters of phytoplankton assemblages are used for assessment of the coastal marine ecosystem.

REFERENCES

1. *Davies R. M., Jensen L. M.* Zooplankton Entrainment at Three Mid-Atlantic Power Plants // *J. Wat. Pollut. Control Fed.* 1974. V. 47. P 2130–2142.
2. *Overnell J.* Inhibition of Marine Algal Photosynthesis by Heavy Metals // *Mar. Biol.* 1976. V. 38 (4). P. 335–342.
3. *Boyer J. N. et al.* Phytoplankton Bloom Status: Chlorophyll a Biomass as an Indicator of Water Quality Condition in the Southern Estuaries of Florida, USA // *Ecological Indicators.* 2009. V. 95. P. 56–67.
4. *Sheue-Duan Lai, Pei-Chung Chen, Hoang-Kao Hsu.* Benthic Algae as Monitors of Heavy Metals in Various Polluted Rivers by Energy Dispersive X-Ray Spectrometer // *J. Environmental Science and Health. Part A — Toxic/Hazardous Substances & Environmental Engineering.* 2003. V. A38 (5). P. 855–866.
5. *Vinogradov A. P.* Elementary Chemical Composition of Marine Organisms. Part 1. Laboratoire de Biogéochimie près l'Acad. des Sci., U.R.S.S., Travaux. 1935. V. 3. P. 63–278 (in Russian).
6. *Leonova G. A. et al.* Concentration of Chemical Elements by Zooplankton of the White Sea // *Marine Biology.* 2013. V. 53 (1). P. 54–70.
7. *Leonova G. A., Bobrov V. A., Bogush A. A.* Estimation of the Geochemical Background and Anthropogenic Pollution of the Plankton by Microelement Composition (by the Example of the Western Siberian Lakes) // *Bioindicators in Monitoring of Freshwater Ecosystems. Abstracts of Intern. Conf. on Environmental Bioindicators, St. Petersburg, Russia, 2006.* P. 91–92.
8. *Martin J. H., Knauer G. A.* The Elemental Composition of Plankton // *Geochim. et Cosmochim. Acta.* 1973. V. 37 (7). P. 1639–1653.
9. *Jensen A., Rystad B., Melson S.* Heavy Metal Tolerance of Marine Phytoplankton. I. The Tolerance of Three Algal Species to Zinc in Coastal Sea Water // *J. Exp. Mar. Biol. Ecol.* 1974. V. 15 (2). P. 145–157.
10. *Hester K., Boyle E.* Water Chemistry Control of Cadmium Content in Recent Benthic Foraminifera // *Nature.* 1982. V. 298. P. 260–262.
11. *Brzezinska A. et al.* Trace Metals in Some Organisms from the Southern Baltic // *Oceanologia.* 1984. No. 18. P. 79–94.
12. *McCormick P. V., Cairns J.* Algae as Indicators of Environmental Change // *J. Appl. Phycology.* 1994. V. 6. P. 509–526.
13. *Vinogradova Z. A., Petkevich T. A.* Chemical Elemental Content of the Black Sea, Azov Sea and Caspian Sea Plankton // *Biochemistry of the Marine Organisms.* 1967. P. 70–82 (in Russian).

14. *Rozhanskaya L. I.* Trace Elements in Plankton and Fishes from the Western Part of the Black Sea // *Ekologiya Morya*. 1983. V. 12. P. 22–30 (in Russian).
15. *Saenko G. N.* Metals and Halogens in Marine Organisms. M: Nauka, 1992. P. 200 (in Russian).
16. *Osadchaya T. S et al.* Organic Carbon and Oil Hydrocarbons in Bottom Sediments of Sevastopol Bay (the Black Sea) // *Marine Ecological J.* 2003. V. 2 (2). P. 94–101.
17. *Frontasyeva M. V.* Neutron Activation Analysis for the Life Sciences // *Particles & Nuclei*. 2011. V. 42 (2). P. 332–378.
18. *Chen G. C., Li C. W., Yang R. S.* Technology of Biological Electron Microscopy. Taiwan: National Science Council, 1991. P. 209–223.
19. Recommended Procedures for Measuring the Productivity of Plankton Standing Stock and Related Oceanic Properties. Washington: NASA, 1969. P. 59.
20. *Parkhomenko A. V. et al.* Results of Hydrochemical Monitoring of Sevastopol Black Sea's Offshore Waters // *J. Coastal Research*. 2003. V. 19 (4). P. 907–911.
21. *Dmitriev A. Yu., Pavlov S. S.* Automated Quantitative Determination of Elements in Samples by Neutron Activation Analysis at the IBR-2 at the LNP JINR // *Particles & Nuclei*. 2013. V. 10 (1). P. 58–64.
22. *Bobrov V. A. et al.* SR XRF Element Analysis of Sea Plankton // *Nucl. Instrum. Meth. Phys. Res. A*. 2005. V. 543. P. 259–265.
23. *Li Yuan Hui.* Distribution Patterns of the Elements in the Ocean: A Synthesis // *Geochim. Cosmochim. Acta*. 1991. V. 55. P. 3223–3240.
24. *David G. Angeler et al.* Does Seston Size Structure Reflect Fish-Mediated Effects on Water Quality in a Degraded Semiarid Wetland // *Environ. Monit. Assess.* 2007. V. 125. P. 9–17; DOI 10.1007/s10661-006-9234-5.
25. *White P. J., Brown P. H.* Plant Nutrition for Sustainable Development and Global Health // *Annals of Botany*. 2010. V. 105 (7). P. 1073–1080.
26. *Mackenzie F. T., Stoffyn M., Wollast R.* Aluminum in Seawater: Control by Biological Activity // *Science*. 1978. V. 199 (4329). P. 680–682.
27. *Morel F. M. M., Reuter J. G., Price N. M.* Iron Nutrition of Phytoplankton and Its Possible Importance in the Ecology of Ocean Regions with High Nutrient and Low Biomass // *Oceanography*. 1991. V. 4. P. 56–61.
28. *Cushing C. E., Rancitelli L. A.* Trace Element Analyses of Columbia River Water and Phytoplankton // *Northwest Science*. 1972. V. 46 (2). P. 115–121.

Received on February 19, 2014.

Корректор *Т. Е. Попеко*

Подписано в печать 18.03.2014.

Формат 60 × 90/16. Бумага офсетная. Печать офсетная.

Усл. печ. л. 1,18. Уч.-изд. л. 1,59. Тираж 225 экз. Заказ № 58217.

Издательский отдел Объединенного института ядерных исследований
141980, г. Дубна, Московская обл., ул. Жолио-Кюри, 6.

E-mail: publish@jinr.ru

www.jinr.ru/publish/